Trends in hybrid cucumber development

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Abstract

Cucumber which belongs to the family Cucurbitaceae, is an important vegetable crop with regard to production value. Its immature fruits are mainly consumed as salad, for making pickles, preparation of cosmetic items, soap and cream. Due to their high-water content, they have refreshing properties. Breeders have vigorously pursued improvements in terms of yield, disease resistance, fruit quality and other economically important traits. This crop exhibits high degree of cross-pollination, wide range of genetic variability in vegetative and fruit characters. A number of studies has been done in relation to genetic improvement, resistance for biotic, abiotic stresses, mutation breeding and on biotechnological aspects. In this background, this review deals with the breeding achievements in cucumber and a special emphasis has been laid on the exploitation of genetic resources for the development hybrids with desirable traits.

Key words: Breeding, cucumber, genetic resources, hybridization and pollination

Introduction

Cucurbits are composed of 118 genera and 825 species and primarily distributed in tropical and subtropical regions of the world (Wang et al. 2007). Globally, watermelon (*Citrullus lanatus*), cucumber (*Cucumis sativus*) and melon (*Cucumis melo*) tops the list of cucurbits in terms of production. The genus *Cucumis* includes 30 wild and cultivated species that are spread throughout the world and has two major species: cucumber and melon. Cucumber (*Cucumis sativus* L., 2n = 2 x = 14) is a widely cultivated and most popular vegetable in the Cucurbitaceae family. Cucumber is well

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known as a fresh market vegetable, primarily grown for its edible tender fruits used as salad and even grown on commercial scale all around the world. Cucumber is a creeping vine which wraps around trellises or other supporting frames with the help of thin and spiraling tendrils. Its fruit is roughly cylindrical, elongated with tapered ends, and the large leaves form a canopy over the fruit. Cucumber contains usually more than 90% water. Sex expression is an important factor which has a positive effect on yield and constitutes a major component of cucumber improvement programs (Serquan et al. 1997).

Origin and Evolution

Cucumber is thought to be indigenous to India. Cucumber has been cultivated in India for over 3000 years and in Eastern Iran and China probably for 2000 years. China is a secondary center of genetic diversification of C. sativus. Cucumber was introduced from India to China, North Africa and South Europe, and from Europe to New World by early travellers and explorers. It was introduced to Tropical Africa by the Portuguese (Staub et al. 1999 and Wang et al. 2007). The wild relative of cucumber C. sativus var. hardwickii is found in Himalayan foothills of India which is fully cross compatible with cucumber. Several accessions of C. sativus var. hardwickii have been collected and it was found that its range of variability fell within that of Cucumis sativus. This fact supports that C. sativus var. hardwickii is either a feral or progenitor of cultivated cucumber and India has been the primary centre of origin for cucumber. The subgenus Cucumis includes Sino-Himalayan species like C. sativus (2n = 2x = 14) and C. hystrix Chakr. (2n = 2x = 24). The wild C. hystrix has unique genetic traits and found only in Yunnan province of Southern China and (Prohens and Nuez 2008). C. sativus has several botanical groups like var. sativus, the cultivated cucumber and var. hardwickii. the wild form. Commercial cucumber, referred to as Cucumis sativus is thought to have originated in the southern Himalayan foothills region of Asia.

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Distribution and Cytogenetics

Cucumis has more than 30 species of which cucumber and melons are economically important origin in India having 2n= 14. Jaffrey (1980) classified the entire genus *Cucumis* in two subgenera:-

- 1) Subgenus Cucumis x=7, 3 or 4, Sino-Himalayan species including *C. sativus* (*C. hystrix, C. muriculatus*).
- 2) Subgenus melo x= 12 species mostly in Tropical and Soth Africa.
- a. Anguria group- Dioecious, monoecious and andromonoecious perennial or annual with yellowish or brown fruit, closely related and partially fertile interspecific hybrids. *C. anguria, C. dispaceus, C. prophetarum, C. myriocarpus, C. sacleuxii.*
- b. Metuliferus group- Monoecious, annual with red spiny fruits, *C. metuliferus*.
- c. Melo group- Monoecious, andromonoecious perennial or annual with smooth fruit. C. melo, C. sagittatu (syn. C. anglolensis), C. dinteri, C. humifructus.
- d. Hirsutus- dioecious perennials with smooth orange fruit, *C. hirsutus*.

The evolution of the genus Cucumis, in particular the relationship between two basic numbers x=7 and x=12 is not clear. Whitaker (1930), Bhaduri and Bose (1947) assumed that species with 2n=24 chromosomes have arisen from species with 2n=14 chromosomes by fragmentation. Trivedi and Roy (1970), however, are of the opinion that by fusion of chromosomes with (sub) terminal centromere species with 14 chromosomes have arisen from species with 24 chromosomes.

Genetic divergence

Genetic diversity is the amount of heritable variability between varieties or populations of organisms. This variability arises due to differences in DNA sequences, biochemical characteristics like isoenzyme properties, physiological properties like resistance to illnesses and growth rate, and morphological characters such as leaf type and flower colour. Selection, mutation, genetic drift and gene flow also affect genetic diversity in different populations by acting on the alleles in these populations. The degree of genetic diversity in C. s. var. sativus is relatively low compared with other cross-fertilized species of Cucumis (Esquinas-Alcazar 1977 and Knerr et al. 1989). Genetic markers (morphological and biochemical) have been employed to characterize the genetic diversity present in the cucumber collection (Knerr et al. 1989 and Meglic et al. 1996).

Pollination and Fertilization

Cucumber is basically monoecious in nature. However, cultivars having gynoecious or predominately female sex expression have been subsequently bred for commercial cultivation. Both male and female flowers are yellow with 5 petals. Male flowers have no pistils but 3 stamens. Female flowers have well developed pistils with three bilobed stigmas, a style and a threechambered ovary but stamens are reduced and nonfunctional. The stigma is receptive throughout the day but most receptive in early morning (Seaton et al 1936). Female flowers produce a higher volume of nectar than male flowers, but sugar concentration is higher in male flowers (Collison 1973). Flowers open in the early morning around 5:30 to 7:00 am and pollination is generally completed by noon. Pistillate flowers are receptive in the morning or upto mid day on the day of opening. Gynoecious genes was more

Table 1: Genetic resources for different traits in cucumber	(Rai and Rai 2006)
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Trait	Promising accession (s)
High yield	Chamba Local-11, EC-27080, Dhaneswar-2, Kheera Bharasati, VRC-11, FLCU-4,
	Patna-3, CH-124, CH-123, CH-136, HAC-71, HAC-31, HAC-26, UHF-Sel, LC-1,
	Ranch-9, VRC-7, Bihar-10, IC-203938, RHRC-1, RHRC-2 and Market Long
Extended shelf life (4 months)	IC-203838, 203839
Early determinate	EC-398030
Gynoecious line	EC-382737, 329300 and 382739
Powdery mildew resistance	Poinsett, Yomaki, Spartan and Saladpi-197088
Watermelon mosaic virus (WMV-1 & WMV-2) resistant	Table green and Surinam
Bacterial wilt resistant	PI-200815, PI-200818, 79-48 and 79-42-24
Multiple disease resistant	EC-320526
Anthracnose resistant	PI-197087
Angular leaf spot resistant	PI-200815, PI-169400, MSU-9402 and GY-14A
Cucumber green mottle mosaic virus	Cucumis anguria
Cucumber mosaic virus	TMG-1, Chinese Long, Chinese Long Green, Shamrock, Wisc-SMR 12, 15, 18, Elem

Trait	Importance	Gene symbol assigned
Bushy	Short internodes	by or bu
Compact	Reduced internodal length	cp
Compact-2	Short internodes	cp-2
Determinate habit	Short vine	de
Determinate habit-2	Short vine	de-2
Female	High degree of pistillate sex expression	F or Acr
Gynoecious	High degree of pistillate sex expression	gy
Trimonoecious	Producing male, female and bisexual flowers	Tr
Andromonoecious	High degree of male expression	m
Multi pistillate	Several pistillate flowers per nodes	mp, mp-2
Parthenocarpy	Sets fruit without pollination	pc
Black or Brown Spine	Fruit quality	B Dominant to white spines
Black Spine-2	22	B-2 Interact with B to produce F ₂ of 15 black: 1 white
Black Spine-3	22	B-3 Interact with B-4 to produce F_2 of 9 black: 7 white
Black Spine-4	"	B-4 Interacts with B-3
Cucurbitacins	Bitter free, lacking cucurbitacin	bi
Dull fruit skin	Fruit colour	D Dull skin of American cultivars dominant to glossy skin of European cultivars
Green mature fruit	Fruit colour	gn
Numerous spines	Fruit quality	ns Few spines is dominant to numerous spines
Male sterile-1,2	Male sterlity	ms-1,ms-2
Bacterial wilt	Resistance to Erwinia tracheiphila	Bw
CMV resistance	Virus resistance	Cmv
Cladosporium resistance	Resistance to scab	Ccu
Downy mildew resistance	Disease resistance	dm-1, dm-2, dm-3
Nematode resistance	Disease resistance	mj
Powdery mildew resistance-1,2,3	Disease resistance	pm-1,2,3
Powdery mildew resistance-h	Resistance expressed by hypocotyl	pm-h
Anthracnose	Disease resistance	Ar
Pseudomonas lachrymans	Disease resistance	psl
Colletotrichum lagenarium	Disease resistance	cla
Corynespora melonis	Disease resistance	Cm
Zucchini yellow mosaic virus	Disease resistance	zymv
Fusarium wilt	Disease resistance	Foc

Table 2: List of genes controlling different traits in cucumber (Navar and More 1998 Ram 1998)

stable under low temperature of 24-25°C and photoperiod upto 12 hr (More and Munger 1986). Light intensity and time of the day influenced anthesis of cucumber more than temperature. The female and male lines are grown in alternate rows in the ratio of 4:1. Cucumbers may also be pollinated by bees.

Some cultivars of cucumber are parthenocarpic *i.e.* formation of seedless fruit without pollination. Pollination for these cultivars degrades the quality. In case of traditional cultivars, male flowers are produced earlier then female flowers, but in almost equal numbers. Newer gynoecious hybrid cultivars produce almost all female flowers. They may have a pollenizer cultivar interplanted, and the number of beehives per unit area is increased, but temperature changes induce male flowers even on these plants, which may be sufficient for pollination to occur. In order to avoid killing off insect pollinators, insecticide applications for insect pests must be done with extra care. Wehner and Kumar (2012) reported that 'NC-Sunshine' is a new monoecious slicing hybrid cucumber with a high percentage of pistillate nodes. Though the percentage of pistillate nodes is high, a pollenizer might be required to maximize pollination

which leads to increase in total and early yield. Golabadi et al. (2019) reported that pollination at 8:00 9:45 AM, led to higher amount of full seed weight in contrast to other times of pollination. Increasing crossed node spacing from 2 3 to 4 5 nodes led to an increase in the number of empty seeds, which was ascribed to the reduced seed production. The highest amount of number of seeds per fruit, seed weigh and number of full seeds were obtained when young male flowers were used.

 Table 3: Main sex types present in cucumber (Singh 2011)

Sex form	Flower type
Hermaphrodite	Bisexual flowers.
Monoecious	Staminate and pistillate flowers.
Androecious	Only staminate flowers.
Gynoecious	Only pistillate flowers.
Andromonoecious	Staminate and hermaphrodite
Gynomonoecious	Pistillate and hermaphrodite
Trimonoecious	Androecious, Gynoecious & Hermaphrodite

Plant micro-environmental factors control expressions of flowering genes of plants of Cucurbitaceae. The ratio of number of female flowers to male flower was higher in favourable condition and less under stress microenvironmental condition. The higher the temperature, the less the moisture content in the soil, within field

capacity to plant wilting point, and the less the inter plant spacing, the higher was the maleness, when the intensity of sunshine and day length remained the same. The expression of female flowering gene was more stress sensitive (Das 2008). Ito and Saito (1957) reported that female flower formation of cucumber is hastened by IAA, NAA and 2,4-D. NAA was reported to be the most effective (Ito and Saito 1956 and Trebitsh et al. 1987). Gibberellin (GA), aminoethoxyvinylglycine (AVG) and silver ions can promote staminate flower development on gynoecious plants (Fuchs et al. 1977, Lower 1978, Den Nijs and Visser 1980, Hunsperger et al. 1983, More and Hunger 1986 and Staub and Crubaugh 1987). Tiedjens (1928) found that pistillate flowering is favored by low light intensity and short photoperiod. An interaction of short days and cool nights favors formation of more 12 pistillate flowers (Ito and Saito 1957, Delia et al. 1982 and Odland and Groff 1962). Kie³kowska (2013) tested the effects of plant growth regulators (PRGs) on the induction of flowering and sex expression in micropropagated cucumbers are presented. The highest number of male flowers (6.0 \pm 0.7 per plant) was produced by cv. Kmicic F1 on the Murashige and Skoog (MS) medium supplemented with 4.0 iM kinetin. The highest number of female flowers (3.1 ± 0.3) was also observed in cv. Kmicic F1 on either control (PRG-free) medium or medium supplemented with 6.4 iM indole-3-acetic acid (IAA).

Interspecific Hybridization

Interspecific hybridization is used to improve crops by transferring specific traits, such as pest and stress resistance, to crops from their wild relatives (Bowley and Taylor 1987). Attempts to introduce valuable traits from wild *Cucumis species* into cultivated cucumber and melon through conventional hybridization have not suceeded because of interspecific cross incompatibilities (Whitaker 1930, Smith and Venkat Ram 1954, Deakin *et al.* 1971 and Fassuliotis and Nelson 1988). Biotechnological techniques such as somatic hybridization have been suggested as possible tools for

overcoming interspecific hybridization barriers in Cucumis (Tang and Punja 1989 and Chatterjee and More 1991). Chen et al. (2002) studied reciprocal differences in interspecific hybridization in *Cucumis*. The fertility of reciprocal plants was greatly different. When C. hystrix was used as female, the tetraploid (2n=38)/diploid (2n=19) plants had higher fruit sets, and even the amphidiploidy plants could produce viable seeds. However, when the cucumber was used as female, both tetraploid/diploid plants were highly sterile and had very low fruit sets. Chen et al. (2005) developed a new pickling cucumber line 7012A through interspecific hybridization of cultivated cucumber Beijingjietou with the Cucumis wild species C. hystrix and subsequent backcrossing of the hybrid to cucumber and selection from the selfed progenies. This line was used to cross with an elite American pickling cucumber line (7011A) to produce F_1 . The results indicated that the F_1 has significant heterosis over its parents in yield and growth vigour. The plants set uniform fruits with good quality and highest yield.

Tak et al. (2016) reported that five best hybrids (L3xT3, L3xT2, L2xT2, L12xT1 and L3xT1) identified based on *per se* performance on pooled basis for total yield per vine. Appreciable heterosis was observed over better and top parent for most of the characters studied. Best three economic hybrid L3 x T3, L7 xT1 and L6 x T2 showing 10.75%, 22.30% and 41.39% economic heterosis for total yield per vine, T.S.S and for fruit length respectively, may be exploited for commercial cultivation.

Mutation Breeding

Mutation is a sudden heritable change in a characteristic of an organism. This definition requires that the change in the characteristic be heritable, but it does not specify the genetic basis of the heritable change. Mutations arise normally in large populations but are usually considered to be rare events. Seeds of cucumber cv. M15 were treated with 0.05% ethylenamine for 21h, and selection

Table 4: Wide-cross attempts between cultivated and wild *Cucumis species*

Cross	Result	Source
C. sagittatus × C. melo	Embryos only	Deakin et al. 1971
C. metuliferus × C. melo	Embryos only	Fassuliotis 1977
C. sativus \times C. melo	Globular stage embryos only	Szczytt and Kubicki 1979
C. metuliferus × C.melo	Fertile F ₁	Norton and Granberry 1980
C. prophetarum × C. melo	Fruit with inviable seeds	Singh and Yadava, 1984
C. zeyheri × C. sativus	Fruit with inviable seeds	Custers and Den Nijs 1986
C. sativus × C. metuliferus	Embryos only	Franken et al. 1988
C. melo \times C. metuliferus	Embryos only	Soria et al. 1990
C. sativus × C. hystrix	Sterile plants (2n and 4n)	Chen et al. 1997
C. hystrix \times C. sativus	Fertile plants (4n)	Chen et al. 1998

for resistance to *Meloidogyne* spp. revealed that 7.6 % of plants were resistant (Udalova and Prikhod'Ko 1985). Swarna Ageti variety of cucumber has been developed with the exploitation of mutation breeding by HARP, Ranchi and it has resistance to powdery mildew. Gamma irradiation of 0.1 kGy stimulated the development of higher number of haploid embryos in cucumber (Faris et al. 1999). Chen et al. (2018) optimized the conditions of EMS mutagenesis on inbred line 406 which included treatment of seeds at 1.5% EMS for 12 h. We obtained a number of mutant lines showing inheritable morphological changes in plant architecture, leaves, floral organs, fruits and other traits through M1, M2 and M3 generations. The F2 segregating populations were constructed and analyzed. We found that a short fruit mutant and a yellow-green fruit peel mutant were both under the control of a single recessive gene, respectively. These results provide valuable germplasm resources for the improvement of cucumber genetics and functional genomic research.

Heterosis Breeding

In cross-pollinated crops, exploitation of heterosis (hybrid vigor) is an important aspect of its improvement. In monoecious crop, the development of F, hybrids is a simple and less costly. The seed requirement per hectare for commercial cultivation would be low and cost effective due to cultivation at wider spacing. Therefore, cucumber offers greater scope for exploitation of hybrid vigour on commercial scale. In cucurbits, high productivity is one of the important aspects of heterosis, but early maturity is equally important especially in cucumber and summer squash, because of the economic dimensions in terms of the price of the early produce. Manifestation of heterosis has attained the highest perfection in cucumber by utilization of gynoecious sex forms, which enabled every flowering node to produce a fruit. Hayes and Jones (1976) reported that first generation crosses in cucumber exhibited high parent heterosis due to increased fruit size and number of fruits per plant. They provide early and higher yield, uniformity in fruit shape, size, colour, better quality and resistance to several disease controlled by dominant genes. Airina et al. (2013) reported that the hybrids EC 709119 \times IC 538155 followed by EC 709119 × IC 527427, EC 709119 × IC 538186 and EC $709119 \times IC 410617$ exhibited high heterobeltiosis for fruit yield and fruits per plant. These hybrids can be advanced for further testing for commercial exploitation of hybrid vigour. Tiwari and Singh (2016) evaluated twenty-four F₁ hybrids derived from 11 diverse cucumber genotypes to study the extent of heterosis and combining ability on earliness and yield characters. Verma & Kumar : Trends in hybrid cucumber development

Appreciable heterosis in desirable direction was found in cross PCUCP-3 \times PCUC-15 for earliness, whereas Cross Kian × PCUC-15 showed maximum number of fruits per plant and fruit vield. Moreover, Simi et al. (2017) observed highest positive heterotic effect for no. of fruits per plant was observed in Modhumoti × Baromashi (20%). The highest heterobeltiosis effect was found in hybrid Himaloy \times Yuvraj (24.5%) followed by Sobujsathi× Khira (11.2 %), Modhumoti × Baromashi (10.0 %). Four crosses exhibited significant positive better parent heterotic effect for this trait and the combination Sobujsathi × Baromashi had the maximum heterosis on yield (47.6%). The maximum heterobeltiosis effect was found in Shila × Khira (27.73 %) followed by Modhumoti \times Hero (15.14%) and Modhumoti × Khira (10%) for fruit yield. Further, Malav et al. (2018) carried out study on heterosis and combining ability for fruit yield and its components in 42 F, hybrids of cucumber obtained from a full diallel involving three gynoecious and four monoecious parents and reported that Hilton × Swarna Sheetal exhibited high heterosis for fruit yield per plant. Sahoo and Singh (2020) used line \times tester mating design to determine the magnitude of heterosis for earliness, yield and yield related traits in cucumber. Appreciable heterosis was observed over better parent and standard parent for most of the characters studied. The best F₁ hybrids reported were PCUCP-4 × PCUC-8 for yield per plant, PCUCP- $3 \times$ PCUC-8 for average fruit weight and PCUCP-1 \times PCUC-8 for node number to first female flower over standard check Pant Khira-1 and these three hybrids may be exploited for commercial cultivation.

Breeding for Multiple Disease Resistance

Development of cucumber hybrids with multiple disease resistance is the most effective and straightforward way of plant protection. The choice of accurate parental lines is the first and foremost step towards successful development of hybrids with desirable traits. Breeding for disease resistance was emphasized in USA with the development of CMV resistant cultivar Shamrock during 1937. In 1955 resistance to scab and CMV were combined in the line Wisconsin SMR-12. Phenomenal success has been accomplished in cucumber by Peterson and his associates (1982) in Wisconsin (USA), where resistant line, WI2757 had been developed carrying resistance to as many as nine diseases (powdery mildew, downy mildew, scab, anthracnose, angular leaf spot, bacterial wilt, target leaf spot, CMV and fusarium wilt). Additional resistances were identified and eventually combined in the lines Sumter with resistances to 7 diseases and Wisconsin 2757 with resistance to 9 diseases. Kushnereva (2008) has developed cucumber

lines and hybrids combining complex resistance to 5 or 6 diseases (downy mildew, powdery mildew, scab, blotch or target spot, Fusarium wilt) with other economically important traits (parthenocarpy, improved fruit quality, etc.). Disease resistant traits are quantitative traits that are controlled by multiple genes, which are generally located in multiple effect quantitative trait loci (QTL). The Gy14 and WI2757 cucumber inbred lines are resistant to multiple diseases and used in the QTL mapping studies for downy mildew, angular leaf spot and anthracnose resistance and further map-based cloning to identify the candidate genes for the resistant loci (Wang et al. 2019). QTL studies have also been carried out by involving WI2757 line for resistances to diseases such as powdery mildew, downy mildew, fusarium wilt, scab and different potyviruses (Wang et al. 2020).

Breeding for Salt and Drought Stress

Sacala et al. (2008) studied effect of water stresses on growth, nitrogen and phosphorus metabolism in cucumber seedlings. Under PEG treatment reduction in cucumber drymass is lesser than in fresh mass, whereas under salt stress decrease in dry weight of cucumber shoots was more pronounced than in fresh mass. NaCl and PEG caused approximately 40% decrease in fresh weight of cucumber shoots (hypocotyl + cotyledons) but there was 45% decrease in dry weight under salt stress and 27% reduction under PEG treatment. Salt stress is more deterimental than sole osmotic stress (PEG treatment). Primary consequence of salinity and drought is osmotic stress, which creates decrease in water availability. Turgor of cucumber seedlings grown under PEG and NaCl was not reduced and water contents in different organs were similar to these in control plants.

Breeding for Low Temperature

Cucumber is one of the more chilling-sensitive crops. It does not grow at temperatures below 16°C and is susceptible to chilling injury, especially at temperatures below 6°C. As a result, in many geographical regions where the crop is planted before the risk of frost has ended (e.g., northern Europe and eastern United States), the cucumber crop is at risk in early spring plantings. Younger seedlings (cotyledons stage) are less sensitive to chilling injury than seedlings at the first true fully expended leaf (Smeets and Wehner 1997). Cultivars with low temperature tolerance are currently unavailable, although differences in response to chilling temperatures were found among the cultivars present on the market (Cabrera et al. 1992). With the development of protected horticulture, cucumber breeding for chilling tolerance has become important. There are just a few literatures on inheritance of cucumber chilling tolerance. Two chilling tolerant lines (Nongdachunguang, Beijing cigua) have been identified and used in genetic study for chilling tolerance. Smeets and Wehner (1997) identified chilling resistance in AR75-79 and 'Chipper', and also developed a method for testing the chilling resistance in cucumber seedlings. Chung et al. (2003) investigated the inheritance of chilling injury in progenies of resistant 'Chipper' and 'AR75-79' crossed with susceptible 'Gy 14' and reported that chilling resistance was maternally inherited. Wadid et al. (2003) reported that PI 267742 showed best mean performance for total yield and related traits under low temperature condition and this parent could be improved and used in producing a low temperature tolerance variety. Klosinska et al. (2013) reported that cucumber cultigens B 5669, PI 390953, and PI 246903 showed low temperature tolerance. Among them B 5669 may become the most desirable to breeders as it exhibits cold germinability combined with good fruit quality traits.

Breeding for Insect Pest Resistance

Cucurbits are highly susceptible to several biotic and abiotic stresses. Simple selection method has been used mostly for developing resistant varieties in cucurbits. The growing of resistant varieties has been a major successful control tactic against the vegetable pests, because of the difficulty of using pesticides on these edible plants. Da Costa and Jones (1971), Kooistra (1971) and Howe et al. (1976) supported this view. In cucumber, single recessive gene bi (bitter free) conferred resistance to different species of cucumber beetles (De Costa and Jones 1971). Munshi et al. (2008) studied genetics of resistance to Cucumber mosaic virus (CMV) in Cucumis sativus var. hardwickii, the wild progenitor of cultivated cucumber. As compared to 31 genotypes of C. sativus var. hardwickii collected from 21 locations in India, IC-277048 (6.33%) was recorded with the lowest mean percent disease intensity (PDI). The results revealed that CMV resistance in C. sativus var. hardwickii was controlled by a single recessive gene. Considering the cross compatibility between C. sativus var. hardwickii and cultivated cucumber, with the help of simple backcross breeding resistance trait can be easily transferred to cultivated species.

Breeding for Disease Resistance

Velkov and Alexandrova (2002) found that the accessions PI 192940, 73-508, Nanchi White Spine, Linia 75-6, Bursa 94-16, Polaris DPM, Linia 61 and Poinsett were suitable sources for combining resistance to powdery mildew with good fruit quality. Velkov et al. (2009) found that three pickling (W 1922, LV 41,

Sumter) and eight salad genotypes (Santo, Long Green, Stono, Polaris S, Sc50, Sagami Hanjioro, Poinset DPM, Taichong Mou Gua) showed combined resistance to both powdery mildew and downy mildew pathogens. Call et al. (2012) screened cucumber for resistance to downy mildew caused by *Pseudoperonospora cubensis* and found PI 605996, PI 330628, and PI 197088 as the most resistant cultigens over all environments. The inheritance of cucumber disease resistance described

- *Pseudoperonospora cubensis* (downy mildew) resistance either by multigenes or a single gene.
- *Sphaerotheca fuliginea* (powdery mildew) resistance by recessive multigenes.
- *Fusarium oxysporum* f.sp. *cucumerinum* (Fusarium wilt) by single dominant or recessive gene.
- *Phytophthora melonis* by resistance by polygenes.
- *Colletotrichum orbiculare* by single dominant gene or recessive gene.
- *Pseudomonas syringae* pv. *lachrymans* (Bacterial angular leaf spot) either by polygene or single recessive gene.
- *Cladosporium cucumerinum* (Scab) by single dominant gene.

Breeding for Nematode Resistance

There are several species of *Meloidogyne* (Root knot nematode) affecting many cucurbits. Efforts to transfer resistance from *C. metuliferus* have not been successful (Fassuliotis 1977, Norton and Granberry 1980). *C. hardwickii* carries resistance to *M. javanica*. Chen *et al.* (2001) carried out study to identify resistance of the wild cucumber species, *Cucumis hystrix* to the root knot nematode and to evaluate the transmission of resistance to progenies of its interspecific hybrids with cucumber (*C. sativus*). Reported that *C. hystrix* was highly resistant and *C. sativus* was highly susceptible to the pest. The resistance was shown to be partially transmitted to the F₁s when the interspecific reciprocal crosses were made, and it was further transmitted to the BCF₁ progeny when the chromosome-doubled F₁s

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were backcrossed as the resistance donor to the cultigens.

Polyploidy Breeding and its Achievements

Cucurbits generally are regarded displaying partial autoand partial allogamy and this phenomenon is often cited as a possible reason for less number of natural polyploids in the family. Cucumber haploids are infertile and do not undergo spontaneous diploidization (Przyborowski and Szczytt 1994). Kubicki (1962) found the optimum duration for the production of tetraploids using cucumber seeds soaked in a 1% and 0.4 to 0.7% solution of colchicine was 6 to 24 hours and 12 to 24 hours, respectively. Induced polyploidy has facilitated gene transfer between some related species when crossed at different ploidy levels (Stoskopf et al. 1993). Autotetraploids of *Cucumis sativus* and *C. metuliferus* were obtained at the highest rate when seeds were immersed in 0.5% colchicine for a period of 6 to 8 hrs. (Walters and Wehner 2002).

In-vitro Breeding

Caglar and Abak (1999) induced haploid cucumber embryos by pollination with irradiated pollen and cultured on E20A medium under aseptic conditions. They found that the regenerative ability of embryos was higher at advanced stages (60% in the first year and 80% in the second year) than at globular stage. These embryos also produced haploid plants rapidly (in 3.5 days). In vitro pollination and fertilization have been used to overcome pre-zygotic (factors hindering effective fertilization) and post-zygotic (barriers occurring during or after syngamy) cross-incompatibility in the genus Cucumis (Ondrej et al. 2002). Germination of isolated pollen grains in culture is a prerequisite for several biotechnological manipulations in cucumber (Vizintin and Bohanec 2004). Crossing barriers in inter-specific hybridization within the genus Cucumis can be overcome through in vitro pollination. Pollen grains were isolated directly from mature male flowers and were cultured with mature ovules. The developing ovules when became enlarged and green were transferred to media supporting embryogenesis. They found caseinhydrolysate to be the

Table 5: Following are the resistant sources identified against different insect-pests

Insects	Resistant sources	References
Striped and spotted cucumber beetles	Nappa 63, Ohimo	Wiseman et al. (1961), Nath (1965)
Cucumber beetle	White Wonder, Ashley Wisconsin 2757	Brett and Sullivan (1970), Peterson et al. (1982)
White fly	Ohio MR 200, PI 220860	De Ponti (1979)
Squash vine borer	National Pickling, Straight Eight	Miller (1956)
Aphid	EC 7050, Extra Early Green Prolific	Lal (1979)
Spider mites	Hybrid Long Pickle	Kooistra (1971)
Vegetable leaf miner	Nappa 63, PI 200815, PI 279465	Brett and Sullivan (1970), Eason and Kennedy (1980)

by Jin'an (2004) as:

Diseases	Resistant sources	References
Anthracnose	PI 175111, 175120, 183308, PI 197087	Barnes and Epps (1952)
Powdery mildew	PI 200815, PI 200818, PI 197088, Natsufushinari, Spartan Salad, Poinsett, Yomaki, WI-2757,	Fanourakis (1984)
Downy mildew	Poinsett, China, PI 197087, Palmetto, Pureto Rico-39	Shimizu et al. (1963)
Scab	Highmoor, Amato and Prosos	Andeweg (1956)

Table 6: Resistant sources identified for different diseases

most beneficial component during in vitro pollination and during development of fertilized ovules. To obtain high yield of potential hybrid embryos, there is a need to optimize in vitro protocol (Skalova et al. 2010). Embryo-rescue and in vitro pollination are suitable in vitro techniques for production of hybrid embryos in *Cucumis* (Navratilova et al. 2011).

 Table 7: List of natural polyploids in Cucumis x=7,12

Species	2n
C. ficifolius	48
C. heptadactylus	48
C. aculeatus	48
C. figarei	72

Genetic Emasculation Mechanism

Genetic emasculation mechanisms have been of practical importance in cucumber (*Cucumis sativus* L.) breeding because it can facilitate F_1 hybrid seed production without hand pollination. Five MS forms are known:

- Gynoecious flowering (G): The G character is under the control of several loci. Usually, G is considered to be a form of sex expression rather than MS because it is one of several sex segregates (Pierce and Wehner 1990).
- Apetalous sterile mutant (*ap*): In *ap*, the corolla of both staminate and pistillate flowers are reduced, and anthers become sepal-like (Grimbly 1980). The pistillate flowers develop to their usual size and, if left unpollinated, produce parthenocarpic fruit.
- Pleiotropic pollen-aborted mutant (*ms-1*): The recessive pleiotropic gene *ms-1* determines MS in which failure of staminate flower anthesis and pollen sterility (PS) varies from 30% to 90% (Shifriss 1950). Grimbly (1980) reported that *ms-1* conditions sterility in which staminate flowers are devoid of pollen, and fertility of pistillate flowers is decreased.
- Aborted male flower type (*ms-2*): In *ms-2*, the MS plants are characterized by abortion of the staminate blossoms. In rare instances when the flowers matured to anthesis, only rudimentary anthers that contained no pollen were present (Barnes 1960, Miller and Quisenberry 1978 and Whelan 1974).

• Closed-flower type: The closed-flower variant is controlled by a recessive gene *cl*. Both staminate and pistillate flowers fail to open at maturity (Groff and Odland 1963).

Except for G, other mechanisms have not been used in hybrid cucumber seed production because their inheritance is determined by nuclear genes and because they are associated with undesirable traits such as missing corolla, malformed ovary, and closed female flowers (Grimbly 1980). Zhang et al. (1994) studied the inheritance and response to chemical and environmental factors of a new male sterile mutant in cucumber and reported that pollen sterile character controlled by a single recessive gene, assigned the tentative designation ps. It was not possible to effect changes in the expression of PS by application of cytokinin, indole acetic acid (IAA) or gibberellin (GA₂), and there were likewise no changes in response to temperature and fertilizer treatment. Unlike gynoecy, which is responsive to some external factors, PS is a stable characteristic.

India is the primary centre of origin for cucumber but the breeding efforts for the improvement of this crop has been rather limited. High yield has always been one of the important breeding objectives but genetic variability for yield is restricted in *Cucumis sativus*. Therefore, our focus will have to shift on *Cucumis sativus* var *hardwickii* which has been successfully utilized in USA for development of breeding lines with multiple lateral branching, sequential fruiting habit and resistance to diseases. Breeding for disease resistance should be emphasized and serious efforts should be made for development of varieties or hybrids having multiple disease resistance through gene pyramiding.

सारांश

खीरा कुकुरबिटेसी परिवार का एक सदस्य है और उत्पादन मूल्य के आधार पर एक महत्वपूर्ण सब्जी फसल है। इसके अपरिपक्व फल मुख्य रूप से सलाद के रूप में, अचार, सौन्दर्य प्रसाधन वस्तुएँ, साबुन और क्रीम बनाने के लिए उपयोग किये जाते हैं। उच्च पानी की मात्रा के कारण फलों में ताजापन बने रहने का गुण पाया जाता हैं। प्रजनकों ने अधिक उपज, रोग प्रतिरोधिता, फल की गुणवत्ता और अन्य आर्थिक रूप से महत्वपूर्ण लक्षणों के संदर्भ में अधिक सुधार किया है। इस फसल में उच्च स्तर पर पर—परागण, वर्धीय अनुवांशिक विविधता और फल आधारित घटकों में आनुवांशिक परिवर्तनशीलता की विस्तष्त श्रष्ंखला प्रदर्शित होती है। आनुवांशिक सुधार के लिए अनेकों अध्ययन जैसे—जैविक व अजैविक प्रतिबलों के प्रति प्रतिरोधिता, उत्परिवर्तन प्रजनन और जैव–तकनीकी पहलूओं को समाहित कर किया गया। इस समीक्षा पृष्ठभूमि में खीरा प्रजनन की उपलब्धियों से संबंधित है और वांछनीय लक्षणों के साथ–साथ संकरों के विकास के लिए आनुवांशिक संसाधनों के उपयोग पर विशेष जोर दिया गया है।

References

- Airina CK, Pradeepkumar T, George TE, Sadhankumar PG and Krishnan S (2013) Heterosis breeding exploiting gynoecy in cucumber (*Cucumis sativus* L.). J Trop Agric 51 (1-2): 144-148.
- Andeweg JM (1956) The breeding of scab-resistant frame cucumbers in the Netherlands. Euphytica 5(2): 185-195.
- Barnes WC (1960). A male sterile cucumber. Proc Amer Soc Hort Sci 77: 415–416.
- Barnes WC and Epps WM (1952) Two types of anthracnose resistance in cucumber. Plant Dis Rep 36: 479-480.
- Bhaduri P and Bose P (1947) Cyto-genetical investigations in some common cucurbits, with special reference to fragmentation of chromosomes as a physical basis of speciation J Genet (1947) 48(2): 237-256.
- Bowley SR and Taylor NL (1987) Introgressive hybridization. In: B.R. Christie (ed.). CRC handbook of plant science in agriculture. vol. 1. CRC Press, Boca Raton, Fla.
- Brett CH and Sullivan MJ (1970) The use of resistant varieties and other cultural practices for control of insects on cucurbits in North Carolina. North Carolina Agr Expt Sta Bulletin 440.
- Cabrera RM, Saltveit Jr. ME and Owens K (1992) Cucumber cultivars differ in their response to chilling temperatures. J Am Soc Hortic Sci 117: 802-807.
- Caglar G and Abak K (1999) The effect of season and irradiation doses on haploid embryo production in cucumber (*Cucumis sativus* L.). Proceeding of the VIth Eucarpia Meeting on Cucurbit Genetics and Breeding, Malaga, Spain 25-30.
- Call AD, Criswell AD, Wehner TC, Ando K and Grumet, R (2012) Resistance of cucumber cultivars to a new strain of cucurbit downy mildew. Hort Sci 47(2) 171-178.
- Chatterjee M and More TA (1991) Interspecific hybridization in *Cucumis* spp. Cucurbit Genet Coop Rpt 14: 69.
- Chen C, Qing-zhi C, San-wen H, Shen-hao W, Xiao-hong L, Xiangyang L, Chen Hui-ming and Tian Y (2018) An EMS mutant library for cucumber. J Integr Agric 17(7): 1612-1619.
- Chen JF, Adelberg JW, Staub JE, Skorupska HT and Rhodes BB (1998) A new synthetic amphidiploid in *Cucumis* from a *C. sativus* \times *C. hystrix* F₁ interspecific hybrid. In: J. McCreight (ed.). Cucurbitaceae Evaluation and enhancement of Cucurbit germplasm. ASHS Press, Alexandria, Va. p 336–339.
- Chen JF, Ling MS and Qian ChT (2001) Identification of Meloidogyne incognita (Kofoid & White) Chitwood resistance in Cucumis hystrix Chakr and the progenies of its interspecific hybrid with cucumber (C. sativus L.). J Nanjing Agricult Univ 24: 21-24.
- Chen JF, Staub JE, Tashiro Y, Ishiki S and Miyazaki S (1997) Successful interspecific hybridization between *Cucumis* sativus L. and *Cucumus hystrix* Chakr. Euphytica 96: 413-419.

- Chen L, Chen Jand Staub J (2005) A new pickling cucumber F_1 hybrid bred from interspecific hybridization. China Vegetables 1: 4-6.
- Chung SM, Staub JE and Fazio G (2003) Inheritance of chilling injury: a maternally inherited trait in cucumber. J Am Soc Hortic Sci 128: 526-530.
- Collison 1973. M.Sc. Thesis. Michigan State, USA.
- Custers JBM and Den Nijs APM (1986) Effects of aminoethoxyvinylglycine (AVG), environment and genotype in overcoming hybridization barriers between *Cucumis* species. Euphytica 35: 639–647.
- Da Costa CP and Jones CM (1971) Resistance in cucumber, *Cucumis sativus* L. to three species of cucumber beetles. Hort Sci 6: 340–342.
- Das D (2008) Sex expressions in cucumber (*Cucumis sativus* L.) and flowering statistics. J Interacademicia 12: 575-576.
- De Ponti OMB (1979) Breeding glabrous cucumber varieties to improve the biological control of the greenhouse whitefly. Cucurbit Genet Coop Rpt 2: 5.
- Deakin JR, Bohn GW and Whitaker TW (1971) Interspecific hybridization in *Cucumis*. Econ Bot 25: 195–211.
- Delia PTV, Peterson CE, and Staub JE (1982) Effect of the duration of short-day treatment on the flowering response of a *Cucumis sativus* var. *hardwickii* (R.) Alef. Line. Cucurbit Genet Coop Rep 5: 2-3.
- Den Nijs APM and Visser DL (1980) Induction of male flowering in gynoecious cucumbers (*Cucumis sativus* L.) by silver ions. Euphytica 29: 273-280.
- Eason G and Kennedy GG (1980) Screening cucumbers for resistance to the vegetable leaf miner. Cucurbit Genet Coop Rpt 3: 5-6.
- Esquinas-Alcazar JT (1977) Alloenzyme variation and relationships in the genus *Cucumis*. Ph.D. Dissertation, Univ. of California, Davis.
- Fanourakis, EN (1984) Screening procedures for powdery mildew resistance in the cucumber. Acta Hortic 287: 147-154.
- Faris NM, Nikolova V and Szczytt KN (1999) The effect of gamma irradiation dose on cucumber (*Cucumis sativus* L.) haploid embryo production. Acta Physiol Plant 21: 391-396.
- Fassuliotis G (1977) Self-fertilization of *Cucumis metuliferus* Naud. and its cross-compatibility with *C. melo* L. J Am Soc Hortic Sci 102: 336–339.
- Fassuliotis G and Nelson BV (1988) Interspecific hybrids of *Cucumis metuliferus* × *C. anguria* obtained through embryo culture and somatic embryogenesis. Euphytica 37: 53–60.
- Franken J, Custers JBM and Bino RJ (1988) Effects of temperature on pollen tube growth and fruit set in reciprocal crosses between *Cucumis sativus* and *C. metuliferus*. Plant Breed 100: 150–153.
- Fuchs E, Atsmon D and Halevy AH (1977) Adventitious Staminate Flower Formation in Gibberellin Treated Gynoecious Cucumber Plants. Plant Cell Physiol 18: 1193-1201.
- Fujieda K nd Akiya R (1962) Inheritance of powdery mildew resistance and spine colour of fruit in cucumber. J Jpn Soc Hort Sci 31: 30-32.
- Golabadi M, Ercisli S and Forough A (2019) Environmental and physiological effects on cross-ability and seed production of greenhouse cucumber. J Seed Sci 41 (2): 134-143.

- Grimbly PE (1980) An apetalous male sterile mutant in cucumber. Cucurbit Genet Coop Rpt 3: 9.
- Groff D and Odland ML (1963) Inheritance of closed-flower in the cucumber. J Hered 54: 191–192.
- Hadley HH and Openshaw SJ (1980) Interspecific and intergeneric hybridization. In: W.R. Fehr & H.H. Hadley (Eds.), Hybridization of crop plants. American Society of Agronomy, Madison, Wis.
- Hayes HK and Jones DF (1976) First generation crosses in cucumber. Republic Conference Agriculture Experiment Station 5: 319-322.
- Howe WL, Sanborn JR and Rhodes AM (1976) Western corn rootworm adult and spotted cucumber beetle associations with cucurbita and cucurbitacins. Environ Entomol 5: 1043–1048.
- Hunsperger MH, Hesel DB and Baker LB (1983) Silver Nitrate Induction of Staminate Flowering in Hermaphroditic pickling Cucumbers. Horti Sci 18: 347-349.
- Ito H and Saito T (1956) Factors Responsible for the Sex Expression of the Japanese Cucumber. III. The Role of Auxin on the Plant Growth and Sex Expression. J Hort Assoc Japan. 25: 101-110
- Ito H and Saito T (1957) Factors Responsible for the Sex Expression of the Japanese Cucumber. (VIII). Effects of Long Day and High Night Temperature Treatment of Short Duration, Accompanied by the Growth Substance Spray Application, on the Sex Expression of Cucumber. J Hort Assoc Japan 26: 209-214.
- Jaffrey C (1980) A review of the Cucurbitaceae. Bot Journ Linn Soc 81: 233–247.
- Jin'an GP (2004) Review on cucumber disease resistance inheritance. China Vegetables (1): 63-66
- Kie³kowska, A. (2013) Sex expression in monoecious cucumbers micropropagated *in vitro*. Biol Plant 57 (4): 725-731.
- Klosinska U, Elzbieta UK, Nowicki M and Wehner TC (2013) Low temperature seed germination of cucumber: Genetic basis of the tolerance trait. J Hort Res 21: 125-130.
- Knerr LD, Staub JE, Holder DJ and May BP (1989) Genetic diversity in *Cucumis sativus* L. assessed by variation at 18 allozyme coding loci. Theor Appl Genet 78: 119–128.
- Kooistra E (1971) Red spider mite tolerance in cucumber. Euphytica 20: 47–50.
- Kubicki B (1962) Polyploidy in muskmelons (*Cucumis melo L.*) and cucumbers (*Cucumis sativus L.*). Genet Pol 3: 161–179.
- Kushnereva V (2008) Breeding of cucumber (*Cucumis sativus*) for resistance to multiple diseases and other traits. Cucurbitaceae 2008, Proceedings of the IXth EUCARPIA meeting on genetics and breeding of Cucurbitaceae, INRA, Avignon (France).
- Lal OP (1979) Relative susceptibility of some cucumber and squash varieties to melon aphid, *Aphis gossypii*. Glov. Indian J Plant Prot 5: 208-210.
- Lower RL, Pharate DM and Horst EK (1978) Effects of Silver Nitrate and Gibberellic Acid on Gynoecious Cucumber. Cucurbit Genet Coop Rep 1: 8-9.
- Malav N, Yadav ML and Maurya IB (2018) Heterosis and combining ability in cucumber (*Cucumis sativus* L.). Int J Chem Stud 6(3): 457-460.

- Meglic V, Serquen F and Staub JE (1996) Genetic diversity in cucumber (*Cucumis sativus* L.): I. A reevaluation of the U.S. germplasm collection. Genet Res Crop Evol 43: 533–546.
- Miller Jr. JC and Quisenberry JE (1978) Inheritance of flower bud abortion in cucumber. Hort Sci 13: 44–45.
- Miller LA (1956) Insecticides and varietal resistance in the control of the squash vine borer, *Melittia cucurbitae* (Horr.) in southwestern Ontario. Can J Agr Sci 36: 309-313.
- More TA and Munger HM (1986) Gynoecious Sex Expression and Stability in Cucumber (*Cucumis sativus* L.). Euphytica 35: 899-903.
- Munshi AD, Panda B, Mandal B, Bisht IS, Rao ES and Kumar R (2008) Genetics of resistance to Cucumber mosaic virus in *Cucumis sativus* var. *hardwickii* R. Alef. Euphytica 164: 501-507.
- Nath (1965) Varietal resistance of cucumber, muskmelon and watermelon to the striped cucumber beetle. Indian J Entomol 26: 304-309.
- Navratilova B, Skalova D, Ondrej V, Kitner M and Lebeda A (2011) Biotechnological methods utilized in *Cucumis* research A review. Hort Sci 38: 150-158.
- Nayar NM and More TA (1998) Cucurbits. Oxford and IBH Publishing Co. Pvt. Ltd., New Delhi, Calcutta.
- Norton JD and Granberry DM (1980) Characteristics of progeny from an interspecific cross of *Cucumis melo* with *C. metuliferus.* J Am Soc Hortic Sci 105: 174–180.
- Odland ML and Groff DW (1962) Linkage of Vine Type and Geotropic Response with Sex Forms in Cucumbers *Cucumis sativus* L. Proc Amer Soc Hort Sci 82:358 - 369.
- Ondrej V, Navratilova B and Lebeda A (2002) In vitro cultivation of *Cucumis sativus* ovules after fertilization. Acta Hortic 588: 339–341.
- Peterson CE, Williams PH, Palmer M and Louward P (1982) Wisconsin 2757 cucumber. Hort Sci 17: 268.
- Pierce LK and Wehner TC (1990) Review of genes and linkage groups in cucumber. Hort Sci 25: 605–615.
- Prohens J and Nuez F (2008) Handbook of Plant breeding: Vegetables I: Asteraceae, Brassicaceae, Chenopodicaceae, and Cucurbitaceae. Valencia, Spain. Universisad Politecnica de Valencia
- Przyborowski J and Szczytt KN (1994) Main factors affecting cucumber (*Cucumis sativus* L) haploid embryo development and haploid characteristics. Plant Breed 112: 70-75.
- Rai N and Rai M (2006) Heterosis Breeding in Vegetable Crops. New India Publishing Agency, New Delhi, pp 317-356.
- Ram HH (1998) Vegetable Breeding- Principles and Practices. Kalyani Publishers, Ludhiana, Punjab.
- Sacala E, Demczuk A, Grzys E and Spiak Z (2008) Effect of salt and water stresses on growth, nitrogen and phosphorus metabolism in *Cucumis sativus* L. seedlings. Acta Soc Bot Pol 77: 23-28.
- Sahoo TR and Singh DK (2020) Exploitation of heterosis in cucumber for earliness, yield and yield contributing traits under protected structure. Int J Chem Stud 8 (1): 918-925.
- Seaton HL, Hustan R and Muncie JH (1936) The production of cucumbers for pickling purposes. Mich Agri Experim Stat Spec Bill, pp 131-273.

- Serquan, FC, Bacher J and Staub JE (1997) Mapping and QTL analysis of horticultural traits in a narrow cross in cucumber (*Cucumis sativus* L.) using random-amplified polymorphic DNA markers. Mol Breed 3: 257-268.
- Shifriss O (1950) Male sterilities and albino seedlings in Cucurbits. J Hered 36: 47–52.
- Shimizu S, Kanazawa K, Kato A, Yokota Y, Koyama T (1963) Studies on the breeding of cucumber for resistance to downy mildew. Part 3. Genetic observations for the resistance to downy mildew and other fruit characters. Bull Hort Res Sta A 2:65–81.
- Simi F, Ivy NA, Saif HB, Akter S and Anik MFA. (2017) Heterosis in cucumber (*Cucumis sativus* L.). Bangladesh J Agr Res 42(4): 731-747.
- Singh AK and Yadava KS (1984) An analysis of interspecific hybrids and phylogenetic implications in *Cucumis* (Cucurbitaceae). Plant Syst Evol 147: 237–252.
- Singh D (2011) Vegetable Science. New Vishal Publications, New Delhi, p 207-271.
- Skalova D, Navratilova B, Ondoej V and Lebeda A (2010) Optimizing culture for in vitro pollination and fertilization in *Cucumis sativus* and *C. melo*. Acta Biol Cracov Bot 52: 111–115.
- Smeets L and Wehner TC (1997) Environmental effects on genetic variation of chilling resistance in cucumber. Euphytica 97: 217–225.
- Smith PG and VenkatRam BR (1954) Interspecific hybridization between muskmelon and cucumber. J Hered 45: 24.
- Soria C, Gomez-Guillamon ML, Esteva J and Nuez F (1990) Ten interspecific crosses in the genus *Cucumis*: A preparatory study to seek crosses resistant to melon yellowing disease. Cucurbit Genet Coop Rpt 13: 31–33.
- Staub JE and Crubaugh L (1987) Use of Silver Thiosulfate as a Potential Tool for Testing Gynoecious Sex Stability in Cucumber (*Cucumis sativus* L.). Cucurbit Genet Coop Rpt 10: 18-20.
- Staub JE, Serquen FC, Horejsi T and Chen JF (1999) Genetic diversity in cucumber (*Cucumis sativus* L.): IV. An evaluation of Chinese germplasm. Genet Resour Crop Ev 46: 297–310.
- Stoskopf NC, Tomes DT and Christie BR (1993) Plant Breeding: Theory and Practice. Westview Press, Inc., Oxford, UK.
- Szczytt KN and Kubicki B (1979) Cross fertilization between cultivated species of genera *Cucumis* L. and *Cucurbita* L. Genet Pol 20: 117–125.
- Szczytt KN, Faris MN, Nikolova V, Trojanowska RM and Malepszy S (1994) Optimization of cucumber (*Cucumis sativus* L.) haploid production and doubling. Proceedings of Cucurbitaceae, Texas, South Padre Island.
- Tak S, Kaushik RA and Nath A (2016) Studies on heterosis in interspecific hybrids of *Cucumis*. The Bioscan 11(4): 3155-3159.
- Tang FA and Punja ZK (1989) Isolation and culture of protoplasts of *Cucumis sativus* and *Cucumis metuliferus* and methods for their fusion. Cucurbit Genet Coop Rpt 12: 29–32.
- Tiedjens VA (1928) Sex ratios in cucumber flowers as affected by different conditions of soil and light. J Agr Res 36: 721-748.

- Tiwari R and Singh DK (2016) Study of heterosis and combining ability for earliness and vegetative traits in cucumber (*Cucumis sativus* L.). J Nat Appl Sci 8 (2): 999-1005.
- Trebitsh T, Rudich J and Riov J (1987) Auxin Biosynthesis of Ethylene and Sex Expression in Cucumber (*Cucumis sativus*). Plant Grow Regul 5: 105-113.
- Trivedi RN and Roy R P (1970) Cytological Studies in Cucumis and Citrullus. Cytologia 35(4) 561-569.
- Udalova VB and Prikhod'Ko VF (1985) ByuJleten' Vsesoyuznogo Instituta Gel' mintologii im. K. I. Skryabina, 41: 67-70.
- Velkov N and Alexandrova M (2002) Sources of resistance to causal agents of powdery mildew on cucumber. Bulg J Agric Sci 8:15-18.
- Velkov N, Neykov ST and Chavdarov P (2009) Resistance in *Cucumis sativus* germplasm to causal agents of powdery mildew and downy mildew. Acta Hortic 229-234.
- Vizintin L and Bohanec B (2004) In vitro manipulation of cucumber (*Cucumis sativus* L.) pollen and microspores: Isolation procedures, viability tests, germination, maturation. Acta Biol Cracov Bot 46: 177–183.
- Wadid MM, Medany MA and Abou-Hadid AF (2003) Diallel analyses for yield and vegetative characteristics in cucumber (*Cucumis sativus* L.) under low temperature conditions. Acta Hortic 598: 279-287.
- Walters SA and Wehner TC (2002) Incompatibility in diploid and tetraploid crosses of *Cucumis sativus* and *Cucumis metuliferus*. Euphytica 128: 371–374.
- Wang Y, Bo K, Gu X, Pan J, Li Y, Chen J, Wen C, Ren Z, Ren H, Chen X, Grumet R and Weng Y (2020) Molecularly tagged genes and quantitative trait loci in cucumber with recommendations for QTL nomenclature. Horticulture Research 7 (3) https://doi.org/10.1038/s41438-019-0226-3.
- Wang Y, Tan J, Wu Z, VandenLangenberg K, Wehner TC, Wen C, Zheng X, Owens K, Thornton A, Bang H H, Hoeft E, Kraan PAG, Suelmann J, Pan J, Weng Y (2019) STAYGREEN, STAY HEALTHY: a loss of susceptibility mutation in the *STAYGREEN* gene provides durable, broad spectrum disease resistances for over 50 years of US cucumber production. New Phytol 221: 415-430.
- Wang YH, Joobeur T, Dean RA and Staub JE (2007) Cucurbits. Genome Mapping and Molecular Breeding in Plants 5: Vegetables, Springer-Verlag, Berlin-Heidelberg, pp 315-329.
- Wehner TC and Kumar R (2012) Requirement for pollenizer in new monoecious hybrid cucumber 'NC-Sunshine'. HortTechnology 22: 191-195.
- Whelan EDP (1974) Linkage of male sterile, glabrate seedling, and determinate plant habit in cucumber. Hort Sci 9: 576–577.
- Whitaker TW (1930) Chromosome number in cultivated cucurbits. Am J Bot 17: 1033–1040.
- Wiseman BR, Hall CV and Painter RH (1961) Interactions among cucurbit varieties and feeding responses of striped and spotted cucumber beetles. Proc Amer Soc Hort Sci 78: 379-384.
- Zhang Q, August CG and James RB (1994) Characterizing a cucumber pollen sterile mutant: Inheritance, allelism, and response to chemical and environmental factors. J Amer Soc Hort Sci 119: 804–807.